

Molecular diagnosis and DNA fingerprinting based on *IS6110* of *Mycobacterium tuberculosis* isolated from patients in Iraq

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Abstract: Polymerase chain reaction assay (PCR) used for the detection of *Mycobacterium tuberculosis* strain and fingerprint study. Ten positive cultures on Lowenstein Jensen (LJ) media were collected from specialized center of chest and respiratory disease\Baghdad .genomic DNA was extracted from all isolates and subjected to genetic characterization by (PCR) methods. two specific primers were used in per analysis for detection *16S rRNA* and insertion sequence *IS6110* where commonly used as a target of Mycobacterium. The result of gel electrophoresis showed that all isolates belong to Mycobacterium, and a sequence showed that 99% identity in sequence of *16S rRNA* of *M. tuberculosis* and there is one nucleotide changes, the isolate submission in gen bank NCBI with accession number (MG03060) and 100 identity in sequence of *IS6110 of M. tuberculosis*. The isolate showed there is Ten polymorphism diversity according to copy number of *IS6110* that have, where 80% of *M.tuberculosis* has large copy number (6_10) and 20% have low copy number (1-5) of isolate. This study showed that *M.tuberculosis* isolated in Iraq belong to two groups. Polymorphism groups depended on copy number of IS6110.

Keywords: Mycobacterium tuberculosis, 16S rRNA, IS6110- DNA fingerprinting.

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Induction:

Tuberculosis (TB) is a devastating disease that has already been present in the human population since prehistoric times. the majority of cases of human TB are caused by *Mycobacterium tuberculosis* (MTB), a species among the genus *Mycobacterium* that are acid-fast, non-motile, slow-growing aerobic bacilli (1, 2, 3)

This disease is one of the most devastating infectious diseases. Globally, about one-third of the world population is believed to be infected with the tuberculosis bacillus and new carriers occur at a rate of one per second (4). Despite advances and developments in diagnosing and treating TB, TB a major health burden around the globe. An estimated 10.4 million new cases and 1.8 million deaths occurred in 2015 according to report of WHO. Iraq is considered among eight high TB burden countries in Eastern Mediterranean Region .In Iraq, according to a recent report of the Ministrv of health (MOH), the incidence rate (45/100.000)inhabitants, (5).

Conventional detection of TB is based on a number of protocols, including microscopic examination of smears stained with the Ziehl-Neelsen stain acid-fast bacilli (AFB) (6). Mycobacterial culture has higher sensitivity and specificity than AFB smear but it takes a long time to verify the results as a long incubation time is required (colonies are rarely visible before 6-8 weeks) (7,8,9)

A large number of different molecular methods based on DNA fingerprints have been developed. Molecular methods have been increasingly used in the diagnosis of TB (10). Nucleic acid amplification based techniques are potentially the most rapid and sensitive methods for identification, detection, and susceptibility testing. These methods can potentially reduce the diagnostic time from weeks to days (11,12,13).

Direct sequencing of the bacterial *16S rRNA* gene) has proven to be a stable and specific marker for mycobacterial identification (14, 15, 16).

Eisenach *et al.* (17, 18, 19) described IS6110 as a repetitive DNA sequence of the M. tuberculosis complex genome, and they used IS6110 PCR to detect M. tuberculosis in clinical samples. Since then, many sensitive and rapid PCR- based assays, most of which use IS6110 as target for mycobacterial DNA amplification, have been developed for the direct detection of M. tuberculosis in a variety of clinical samples obtained from adults [20.21]. We use developed a rapid PCR assay that specifically types M. tuberculosis isolates by generating distinct DNA band patterns or fingerprints. This assay utilizes sequences in IS6110, which occurs as

repeated copies in the chromosomes of M. tuberculosis most isolates (22.23.24). The aim of this study using molecular methods for identification of M.tuberculosis isolates from Iraqi patients in Baghdad. And the present study was undertaken to determine the suitability of IS6110 element detection by PCR for the early diagnosis of new TB cases. This assav also has considerable power to differentiate *M.tuberculosis* strains and is highly reproducible by fingerprints.

Materials and Methods:

10 isolates on LJ media previously patient from attended isolated chest specialized center of and disease/Baghdad. respiratory DNA extraction was carried out according to manufactures instruction of Genotype MTBDR pluse kit.

DNA Concentration and Quantification:

Quantity of DNA was confirmed using the NanoDrop ND-1000 spectrophotometer.

PCR for detection of *Mycobacterium tuberculosis*:

Two primers were used in the PCR experiment for the detection of *M. tuberculosis*, and Primer used for fingerprinting, Table(1).

Primers name	Sequences(5-3)	Size	Ref
16S rRNA(f)	ACGGTGGGTACTAGGTGTGGGTTTC	542 bp	25
16S rRNA(R)	TCTGCGATTACTAGCGACTCCGACTTCA	542 Op	23
INS-1	CGTGAGGGCATCGAGGTGGC	245	26
INS-2	AAACAGTGGCTGCGGATGCG	bp	20
F-1	CGCCAGAGACCAGCCGCC		27
F-2	CGCCAGAGACCAGCCGCC		27

Table (1): sequence of primer used for pcr amplification in this study

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PCR mixture was prepared with 25 μ l of Green Master Mix 2x (promega), 17 μ l of nuclease free water, 2 μ l of each primer at 10 pmol/ μ l, and 4 μ l of DNA (equaling 25 to 250 ng). Thermocycling conditions of 95°C (initial denaturation) for 5 minutes, followed by 25 cycles of denaturation at 94°C for 1 minute , and annealing at 60°C for 1 minute and 72°C for 1 minute, then the final extension step at 72 °C for 10 minutes concluded the reaction program.

Products $(10\mu l)$ and DNA molecular weight marker were electrophoresed on 1% Agarose gel with ethidium bromide staining to verify the size of the amplicon, the remaining of

PCR products of 16S rRNA and insertion sequence IS6110 were sequenced at (NICEM-USA, Apparatus: Applied Biosystem).

Results and Discussion:

Ten positive cultures were subject to molecular diagnosis by PCR, the genome of isolates was extracted and purified and The concentration and purity of DNA were measured by nanodrop and the concentration of DNA was ranged between 100 to 420 ng/ μ l and the purity of DNA (A260/A280) ranged between 1.4 to 1.8

Molecular Identification of *M. tuberculosis* isolates:

Molecular identification of M. tuberculosis based on the using of partial 16S rRNA to prove the extracted DNA was belonged to Mycobacterium genus, the product of amplification 542 bp occupying (1472650-1473192), the results are shown that all isolate have 16S rRNA (Figure 1). The amplification of partial 16S rRNA gene was used to confirm the presence of Mycobacterial DNA in the sample (28) The 16S rRNA sequence is an appealing target for the purpose of molecular identification; it is stable property and it contains nucleic acid information allowing the identification of mycobacteria at the genus level as well as the rapid recognition of previously undescribed mycobacterial pathogen (29,30,31).



Figure (1): Gel electrophoresis for PCR product of 16SrRNA 543bp using (1.5%) agarose for 90 minutes at 60 volt.

- 1. M 100 bp ladder.
- 2. c- Negative isolates.
- 3. P- Positive isolates.
- 4. (1-10) PCR product of positive isolates from patients

Presence of insertion sequence *IS6110* in isolates of *M. tuberculosis* complex:

In this study, insertion sequence IS6110 was used to differeniate M. tuberculosis complex from the other of М. tuberculosis subspecies, the presence of IS6110 in M. tuberculosis complex collected from different parts of Baghdad and governorates was tested by PCR using specific primer for *IS6110*. The results showed that all isolate contained *IS6110*. These isolates tested for the presence of the sequence contain the IS6110 suggesting that this sequence is reliable source for the detection and discrimination of *M*. tuberculosis isolated in NRL in Baghdad. (Figure

2). The results indicated that all strains of M. tuberculosis carriers IS6110. Thus it is possible to conclude that IS6110- based diagnosis is practical regardless of the possible encountering of IS6110- free strains. Other studies (32) have confirmed that IS6110 can be used for both DNA fingerprinting of *M. tuberculosis* and for diagnosis of tuberculosis disease specifically. The method is more accurate and faster than conventional method for TB diagnosis. Early diagnosis of *M*. tuberculosis disease is crucial in initiating treatment and interrupting strain transmission. the Rapid diagnosis will prevent the development of drug resistant M. tuberculosis bacteria.



Figure (2): Gel electrophoresis for PCR product of *IS6110* 245pb using (1.5%) agarose for 90 minutes at 60 volt.

- 1. M. 100 bp ladder.
- 2. c- Negative isolates.
- 3. P- Positive isolates.
- 4. (1-10) PCR product of positive isolates from patients

Sequence of the *16SrRNA* gene and the *IS6110* insertion sequences element:

Sequencing of the *16s rRNA* gene and the *IS6110* gene for M. tuberculosis were analyzed using the genetic analysis tool and compared with the sequencing of the same gene in the NCBI to confirm the molecular diagnosis of Congenital PCR and differentiation *M.tuberculosis* from other types of Mycobacterium, The results of the current study showed that the identity ratio of *16s rRNA* gene was 99% where the guanin base was replaced with the adenin base in site 1273168 from the genome (figure 3) this sequence of isolation deposited in NCBI with Accesssion number (MG030630) (figure 4) . and the identity ratio of the *IS6110* was 100% (figure5).

Mycobacterium tuberculosis H	37Rv, complete genor	ne
Sequence ID: gi 448814763 NC_0	000962.3 Length: 4411	532 Number of Matches: 1

Range	e 1: 1472	719 to 1473180 🤆	GenBank Graphics	,	🛚 Next Match 🔺 Pr
Score		Expect	Identities	Gaps	Strand
848 b	its(459)	0.0	461/462(99%)	0/462(0%)	Plus/Plus
Query	1	GCCTGGGGAGTACGGCC	GCAAGGCTAAAACTCAAAGGAA	TTGACGGGGGGCCCGCACA	AGC 60
Sbjct	1472719	GCCTGGGGAGTACGGCC	CGCAAGGCTAAAACTCAAAGGAA	TTGACGGGGGGCCCGCACA	AGC 1472778
Query	61	GGCGGAGCATGTGGATT	TAATTCGATGCAACGCGAAGAAC	CTTACCTGGGTTTGACAT	GCA 120
Sbjct	1472779	GGCGGAGCATGTGGATT	TAATTCGATGCAACGCGAAGAAC	CTTACCTGGGTTTGACAT	GCA 1472838
Query	121	CAGGACGCGTCTAGAGA	ATAGGCGTTCCCTTGTGGCCTGT	GTGCAGGTGGTGCATGG	TGT 180
Sbjct	1472839	CAGGACGCGTCTAGAGA	the second contraction of the second s	dtocadottoctocatood	tgt 1472898
Query	181	CGTCAGCTCGTGTCGTG	GAGATGTTGGGTTAAGTCCCGCAA	ACGAGCGCAACCCTTGTC	TCA 240
Sbjct	1472899	cetcaectcetetcete	GAGATGTTGGGTTAAGTCCCGCA	ACGAGCGCAACCCTTGTC	TCA 1472958
Query	241	TGTTGCCAGCACGTAAT	rggtgggggactcgtgagagactg	CCGGGGTCAACTCGGAGG	AAG 300
Sbjct	1472959	téttéccaécacétaat	rggtggggactcgtgagagactg	cceeeetcaactceeaee	AAG 1473018
Query	301	GTGGGGATGACGTCAAG	GTCATCATGCCCCTTATGTCCAG	GGCTTCACACATGCTACA	ATG 360
Sbjct	1473019	GTGGGGATGACGTCAAG	STCATCATGCCCCTTATGTCCAG	GGCTTCACACATGCTACA	ATG 1473078
Query	361	GCCGGTACAAAGGGCTG	GCGATGCCGCGAGGTTAAGCGAA	TCCTTAAAAGCCGGTCTC	AGT 420
Sbjct	1473079	GCCGGTACAAAGGGCTG	scgatgccgcgaggttaagcgaa	tccttaaaagccggtctd	ÁGT 1473138
Query	421	TCGGATCGGGGTCTGCA	ACTCGACCCCGTTAAGTCGGAG	TCG 462	
Sbjct	1473139	TCGGATCGGGGTCTGCA	ACTCGACCCCGTGAAGTCGGAG	TCG 1473180	

Figure (3): Result of alignment of 16S rRNA according to NCBI data.

LOCUS	Seq1		451	bp	DNA	linear	BCT	04-OCT-201
DEFINITION	DNA, parti	ial.						
ACCESSION	Seq1							
VERSION								
KEYWORDS								
SOURCE	Mycobacte	erium tubero	culosis					
ORGANI SM	Mycobacte	erium tubero	culosis					
	Bacteria;	Actinobact	teria; Co:	ryneba	cterial	es; Mycoba	acter	Laceae;
	Mycobacte	erium; Mycol	pacterium	tuber	culosis	complex.		
REFERENCE	1 (bases	1 to 451)						
AUTHORS	Mohammed,	k.M.						
TITLE	Mycobacte	erium tubero	culosis					
JOURNAL.	Unpublish	led						
REFERENCE	2 (bases	3 1 to 451)						
AUTHORS	Mohammed,	k.M.						
TITLE	Direct Su	ubmission						
JOURNAL.		1 (04-OCT-20				, Diyala ı	mive	rsity,
		ıla, diyala,						
COMMENT	Bankit Co	mment: ALT	EMAIL: wal	hj_dna	@yahoo.	com.		
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		ly-Data-STAI						
		ng Technolog		ger di	deoxy s	equencing		
	##Assemb]	ly-Data-END						
FEATURES		Location/Q	alifiers					
Source		1451						
		/organism=			tubercu	losis"		
		/mol_type='		DNA"				
		/strain="M		_	_			
		/isolation						
		/host="Home						
		/culture_co			nstien :	jennies me	edia"	
		/db_xref="t /clone="165						
		/country="1		2				
		/collection						
		/collected			ankhi W			
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gene		<1>451	rearea ba	JUCI IG	r source	-1		
gene		/gene="165	TDNA T					
rRNA		<1451						
		/gene="165	TRNA "					
		/product="1		mal R	NA"			
BASE COUNT	101 a		144 g	95				
ORIGIN								
1 a	corcorcaa	ggctaaaact	caaaggaa	tt gac	aaaaacc	cgcacaago	a ac	rgagcatg
		cgatgcaacg						
		cgttcccttg						
181 g	tcgtgagat	gttgggttaa	gtcccgca	ac gag	cgcaace	cttgtctca	at gti	tgecagea
241 c	gtaatggtg	gggactcgtg	agagactg	ce ggg	gtcaact	cggaggaaq	ag tag	ggatgac
301 g	tcaagtcat	catgcccctt	atgtccag	gg ctt	cacacat	gctacaato	g cc	ggtacaaa
361 g	ggetgegat	gccgcgaggt	taagcgaa	te ett	aaaagcc	ggtctcagt	tt cg	gategggg
421 t	ctgcaactc	gaccccgtta	agtcgaag	te g				
11								

Figure (4): Accession number of isolate (MG030630)

Mycobacterium tuberculosis H371W, complete genome		
Sequence ID: <u>NZ_CP009480.1</u> Length: 4396119 Number of Matches: 16		
Range 1: 888085 to 888267 GenBank Graphics	▼	Nex

Mycobactorium tuborculosis H37Ry, complete genome

Score	Expect	Identities	Gaps	Strand
339 bits(183)	1e-91	183/183(100%)	0/183(0%)	Plus/Plus
)uery 1	CAAACTCGGCCTGTC	CGGGACCACCCGCGGCAAAGCC	CGCAGGACCACGATCGCT	GATCC 60
5bjct 888085	CAAACTCGGCCTGT	CCGGGACCACCCGCGCCAAAGCC	CGCAGGACCACGATCGCT	GATCC 888144
uery 61	GGCCACAGCCCGTC	CCGCCGATCTCGTCCAGCGCCGC	TTCGGACCACCAGCACCI	AACCG 120
5bjct 888145	GGCCACAGCCCGTC	CGCCGATCTCGTCCAGCGCCGC	TTCGGACCACCAGCACCT	TAACCG 888204
Query 121	GCTGTGGGGTAGCAG4	асстсасстатототсоасстоо	GCAGGGTTCGCCTACGTG	GCCTT 180
5bjct 888205	GCTGTGGGTAGCAG	ACCTCACCTATGTGTCGACCTGG	GCAGGGTTCGCCTACGT	GCCTT 888264
uery 181)	TGT 183			
bjct 888265	tgt 888267			

Figure (5): Result of alignment of IS6110 according to NCBI data

PCR Fingerprinting:

molecular The development methods to genetically differentiate M. tuberculosis strains provide a useful epidemiological tool. first The described method for differentiating М. tuberculosis isolates used mycobacteriophages or enzymatic characterization(33) Although useful for these studies of specific outbreaks these techniques very had low discriminatory power (34). DNA fingerprinting of M. tuberculosis isolates has increasingly been used in epidemiological studies. DNA was compared visually for similarity based on presence or absence of bands and variation in bands intensity were taken

to constitute strains differences (35). Our study presents a precise picture of the epidemiology of TB in different regions in Iraq by applying both molecular and classical epidemiological methods. Figure (6) showed fingerprints patterns on agarose gel. The number of copies was distributed within a range of 1-10 copies. These can be classified into two groups, group A, the isolates that have less than 6 copies of IS6110 and group B, and the isolates that have (more than 6 copies of IS6110 in their chromosomal DNA (36) Of 10 M. tuberculosis isolates (20%) belong to group A and (80%) belong to group B.



Figure (6) PCR fingerprinting (agarose 1.5%) of *M.tuberculosis* strains based on the number of IS6110 copies:

- 1. M 100 bp ladder.
- 2. c- Negative isolates.
- 3. P- Positive isolates.
- 4. (1-10) PCR product of positive isolates from patients.

The result show there was clear diversity banding among the ten isolates. Identification and differentiation of strains М. of tuberculosis by molecular technique of targeted insertion elements has provided a better understanding of the epidemiology of infection (37). In the present study different IS6110-based patterns were observed for Mycobacterium tuberculosis isolates from different regions of Iraq. suggesting differences in copy number and genomic location of the element IS6110.the result indicates that there was substantial diversity of IS6110 banding among isolates of М. tuberculosis in Iraq. This may refer to low degree of active transmission of tuberculosis among patients. Cases examined may have arisen from reactivation of previous infection. This study has demonstrated that the majority of isolates have multiple IS6110 copies (80%). These findings indicate that *M. tuberculosis* strains isolated in Iraq are similar to those found in Iraq and other countries.

References:

- 1- Kethireddy, S.; Nalgonda, D. and Pradesh, A. (2010). Tuberculosis-A Review of Clinical Features, Differential Diagnosis and Treatments available. IJPT. 2 (2):206-244.
- 2- Reddington, K.; Zumla, A.; Bates, M.; van Soolingen, D.; Niemann, S.; Barry, T. and O, Grady, J. (2012). SeekTB, a Two-Stage Multiplex Real-Time-PCR-Based Method for Differentiation of the *Mycobacterium tuberculosis* Complex. Journal of Clinical Microbiology. 50(7):2203–2206.
- Alexander, K.; Laver, P.; Michel, A.; Williams, M.; van Helden, P. and Warren, R. (2010). Novel Mycobacterium tuberculosis complex pathogen, M. mungi. Emerg. Infect. Dis. 16(8): 1296-1299.

- **4-** World Health Organization, (2007). Tuberculosis infection and transmission fact sheet 104.
- MOH-NTP (2013). Epidemiological Situation in Iraq, Fifth National Conference of tuberculosis/Al- Basra. Ministry of health of Iraq-Notional TB programme TB.
- 6- Miller, N.; Cleary, T.; Kraus, G.; Young, A.; Spruill, G. and Hnatyszyn, H. (2002). Rapid and Specific Detection of Mycobacterium tuberculosis from Acid-Fast Bacillus Smear-Positive Respiratory Specimens and BacT/ALERT MP Culture Bottles by Using Fluorogenic Probes and Real-Time PCR. J. Clin. Microbiol. 40 (11): 4143-4147.
- 7- Bang, H.; Choi, T. and Shin, J. (2011). Comparison of Ogawa Media, BACTEC MGIT 960 System and TB/NTM Real-Time PCR for Detecting Mycobacterium Species. Tuberc Respir Dis. 71:249-253.
- 8- Manabe, Y.; Zawedde-Muyanja, S.; Burnett, S.; Mugabe, F.; Naikoba, S. and Coutinho, A. (2015). Rapid Improvement in Passive TB Case Detection and TB Outcomes Treatment Following Implementation of a Bundled Laboratory Diagnostic On-Site and Training Intervention Targeting Mid-Level Providers. Oxford University Press on behalf of The Infectious Diseases Society of America.
- 9- Hung, N.; Sy, D.; Anthony, R.; Cobelens, F. and van Soolingen, D. (2007). Fluorescence microscopy for tuberculosis diagnosis. Lancet Infect Dis 7: 238-239.
- 10- Nakajima, C.; Rahim, Z.; Fukushima, Y.; Sugawara, I.; van der Zanden, A.; Tamaru, A. and Suzuki, Y. (2010). Identification of Mycobacterium tuberculosis clinical isolates in Bangladesh by a species distinguishable multiplex PCR. BMC Infectious Diseases. 10: 118.
- 11- Ruiz, M.; Torres, M.; Llanos, A.; Arroyo, A.; Palomares, J. and Aznar, J. (2004). Direct Detection of Rifampin- and Isoniazid-Resistant Mycobacterium tuberculosis in Auramine-Rhodamine-Positive Sputum Specimens by Real-Time PCR. J.Clin. Microbiol., 42(4): 1585–1589.
- 12- Suffys, P.; Palomino, C.; Cardoso-Leao, S.; *et al.* (2000). Evaluation of the polymerase reaction for the detection of Mycobacterium tuberculosis. Int. J. Tuberc Lung Dis; 4: 179–183.

- Baker, C.; Smith, J. and Cowan, D.A. (2003). Review and re-analysis of domainspecific 16S primers. J. Microbiol Meth; 55: 541–555.
- 14- Bottger, E.C. (1989). Rapid determination of bacterial ribosomal RNA sequences by direct sequencing of enzymatically amplified DNA. FEMS Microbiol Lett; 53: 171–176.
- 15- Sontakke, S.; Cadenas, M.B.; Maggi, R.G.; *et al.* (2009). Use of broad range16S rDNA PCR in clinical microbiology. J Microbiol Meth; 76: 217–225.
- 16- Kirschner, P.A.; Meier, and E. C. Bo⁻⁻ttger. (1993). Genotypic identification and detection of mycobacteria: facing novel and uncultured pathogens, 173–190. In D. H. Persing, F. Tenover, T. J. White, and T. F. Smith (ed.), Diagnostic molecular microbiology. American Society of Microbiology, Washington, D.C.
- 17- Kirschner, P.; Springer, B.; Vogel, U.; Meier, A.; Wrede, A.; Kiekenbeck, M.; *et al.* (1993). Genotypic identification of mycobacteria by nucleic acid sequence determination: report of a 2-year experience in a clinical laboratory. J. Clin. Microbiol. 31:2882–2889.
- 18- Springer, B.; Stockman, L.; Teschner, K.; Roberts, G.D. and Bo⁻ttger, E.C. (1996). Two-laboratory collaborative study on identification of mycobacteria: molecular versus phenotypic methods. J. Clin. Microbiol. 34:296–303.
- 19- Van Embden, J.D.; Cave, M.D.; Crawford, J.T.; Dale, J.W.; Eisenach, K.D.; Gicquel, B.; *et al.* (1993). Strain identification of M. tuberculosis by DNA fingerprinting: recommendations for a standardized methodology. J. Clin. Microbiol. 31: 406-409.
- 20- van Soolingen, D.; Hermans, P.W.; De Haas, P.E.; Soll, D.R. and van Embden. J.D. (1991). Occurrence and stability of insertion sequences in Mycobacterium tuberculosis complex strains: evaluation of an insertion sequence-dependent DNA polymorphism as a tool in the epidemiology of tuberculosis. J. Clin. Microbiol. 29:2578–2586.
- **21-** Dalovisio, J.R.; Montenegro-James, S.; Kemmerly, S.A.; *et al.* (1996). Comparison of the amplified Mycobacterium tuberculosis (MTB) direct test, Amplicor MTB PCR, and IS6110-PCR for detection

of MTB in respiratory specimens. Clin. Infect. Dis.; 23: 1099–1106.

- 22- Suffys, P.; Palomino, J.C.; Cardoso-Leao, S.; *et al.* (2000). Evaluation of the polymerase reaction for the detection of Mycobacterium tuberculosis. Int J Tuberc Lung Dis., 4: 179–183.
- 23- McAdam, R.A.; Hermans, P.W.; van Soolingen, D.; Zainuddin, Z.F.; Catty, D.; van Embden, J.D., *et al.*, (1990). Characterization of a *Mycobacterium tuberculosis* insertion sequence belonging to the IS3 family. Mol. Microbiol, 4: 1607-1613.
- 24- Thierry, D.; Cave, M.D.; Eisenach, K.D.; Crawford, J.T.; Bates, J.H.; Gicquel, B.; and Guesdon. J. L. (1990). IS6110, an ISlike element of Mycobacterium tuberculosis complex. Nucleic Acids Res. 18: 188.
- 25- Richard, C.; Luiz, C.; Oliveira, L. and John L. (2003). PCR-Based Method To Differentiate the Subspecies of the Mycobacterium tuberculosis Complex on the Basis of Genomic Deletions. Journal Of Clinical Microbiology, 41: 1637–1650.
- 26- Han, X.; Pham, A.; Tarrand, J.; Sood, P.; and Luthra, R. (2002). Rapid and Accurate Identification of Mycobacteria by Sequencing Hypervariable Regions of the 16S Ribosomal RNA Gene. American Journal of Clinical Pathology, 118: 796-801.
- 27- Huard, R.C.; Lazzarini, LCO; Butler, W.; van Soolingen, D. and Ho, J.L. (2003). PCR-based method to differentiate the subspecies of the Mycobacterium tuberculosis complex on the basis of genomic deletions. J. Clin. Microbiol. 41(4): 1637-1650.
- **28-** Vasconcellos, S.E.; Huard, R.; Niemann, S.; Kremer, K.; Santos, A.R.; Suffys, P.N. *et al.* (2010). Distinct genotypic profiles of the two major clades of Mycobacterium africanum BMC Infectious Diseases. 10: 80.
- **29-** Willi, A.G.; Stuart, M. and Frances, A.W. (1999). Specificity of IS6110-based DNA fingerprinting and diagnostic techniques for Mycobacterium tuberculosis complex. J. Clini. Micro. 37(4): 1224-1226.
- **30-** Bates, J.H. and Fitzhugh, J.K. (1967). Subdivision of the species M. tuberculosis by mycobacteriophage typing. Am. Rev. Resp. Dis. 96: 7-10.

- **31-** Roman, M.C. and Sicilia, M.J. (1984). Preliminary investigation of Mycobacterium tuberculosis biovars. J. Clin. Microbiol. 20:1015–1016.
- **32-** Raleigh, J.W. and Wichelhausen, R. (1973). Exogenous reinfection with Mycobacterium tuberculosis confirmed by phage typing. Am. Rev. Respir. Dis.108: 639-642.
- 33- Neimark, M.; Mirza B. A. and Carlton, S. (1996). Direct identification & Typing of Mycobacterium tuberculosis by PCR. J. Clin. Microbiol. Oct., 2454-2459.
- 34- Yang, Z.H.; De Haas, P.E.; Wachmann, C.H.; Van Soolingen, D.; Van Embden, J.D. and Andersen, A.B. (1995a). Molecular epidemiology of tuberculosis in Denmark in 1992. J. Clin. Microbiol. 33: 2077-2081.
- 35- Van Soolingen, D.; Hermans, P.W.; De Haas, P.E.; Soll, D.R. and Van Embden, J.D. (1991). Occurrence and stability of insertion sequences in Mycobacteriu tuberculosis complex strains: evaluation of an insertion sequence-dependent DNA polymorphism as a tool in the epidemiology of tuberculosis. J. Clin. Microbiol. 29: 2578-2586.