

Staphylococcus aureus Incidence in Some Patients with a Topic Dermatitis in Baghdad City

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Abstract: This study was planned to evaluate the incidence of *Staphylococcus aureus* in cases of atopic dermatitis in Iraq. This study included taking 150 samples of Atopic Dermatitis (AD) instances in total. Patients Suspected infected with *Staphylococcus aureus*, all isolates were subjected to primary identification tests by using various methods (cultural characteristic, gram staining, biochemical tests, vitek2 system, and molecular methods (PCR). Only 54 sample (36% isolates) were given the typical biochemical tests and characteristics of morphology that is specific to *staphylococcus aureus*, while the other 96 clinical (64 % isolates) belong to other species of staphylococci or other pathogenic bacteria and fungi. As these (54) samples contained the (*mecA*) gene, which is the diagnostic gene for the *staph aureus* bacteria in this study. It was concluded *Staphylococcus aureus* infection is positively correlated with atopic dermatitis cases.

Key words: Staphylococcus aureus, atopic dermatitis.

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Introduction

Atopic dermatitis (AD) is a chronic inflammatory disease causing intense pruritus, and with typical clinical features. eczematous dermatitis 10-20% of children and affecting regarded as a major cause for morbidity, since patients with AD have a higher susceptibility for microbial colonization and an increased risk of skin infections (1).Numerous investigations and studies have shown that in addition to bacteria, particularly S. aureus and fungi may also be significant in aggravating factors AD(2). Staphylococcus aureus infection was also found with prevalence of about 1-3% in adults, present in 80-100% of skin from atopic patients and is related to worsening of the disease by the action of enterotoxins (3).

The role of bacteria in AD is largely unknown, however, bacterial flora analysis of the skin has revealed its involvement in clinical conditions. On the skin of children with AD, it has been reported that the diversity of the bacterial flora of the skin decreases in the exacerbation phase, and the proportion of *S. aureus* increases (4).

In addition, eradication or reduction of *S. aureus* colonization on AD patients has demonstrated a positive correlation with decreased severity of atopic eczema (5). Our study planned to detect the spread of *S. aureus* from skin lesions of AD patients.

Materials and methods Samples collection

This study included a total of one hundred and fifty cotton swab samples

collected from patients of both sexes and of different ages with atopic dermatitis. The samples were collected from Al-Zafrania Hospital and private clinics in different areas of Baghdad City during the period from November 2021 to March 2022. The samples were placed in a transport medium and transferred to Al-Madaen General purpose Hospital for the of transplanting it in the laboratory for a period not exceeding 24 hours only. Followed by bacteriological isolation and identification of S. aureus.

Isolation and identification of *S. aureus*

The collection of skin lesion specimens was by the end of wetting sterilized cotton swab with the lesion or discharge of fluid from blisters scrape tissue, then placed in a sterile test tube containing sterilized 2 ml of transport media . All skin swabs were inoculated on 5% human blood agar and mannitol salt agar plates. After incubation at 37Co for 18-24 hr. cultural and morphological features of the colonies were evaluated. These S. aureus isolates were further investigated by growth on chromagar specific for the identification and isolation of S. aureus.

All isolates were primarily examined by gram stain and by biochemical examinations, according to (Prescott) (6).

• Coagulase test (7): То test coagulase enzyme as bound or free coagulase, suspension 0.5ml of bacteria colony is added to 0.5ml plasma of human have been prepared and incubated in human 37C, then the plasma inoculated periodically examine the composition of fibrin formation of coagulase clot within four hours, and this is interpreted as

a positive result indicates on *Staphylococcus aureus* strains. The absence of a blood clot after 24 hours incubation is the negative result.

- Catalase test (8): This test was used for the detection of catalase enzyme, differentiate Streptococci to (Catalase -ve) from Staphylococci (Catalase +ve). The exposed to hydrogen peroxide, bacteria catalase positive conversion of peroxide to water and oxygen gas, this test was performed by a capillary tube containing hydrogen peroxide solution was carefully dipped into a single colony, and when catalase was represented, oxygen gas was released and the bubbles observed in the capillary tube.
- Oxidase test (9): A piece of filter paper placed in a clean Petri dish and 2-3 drops of Oxidase reagent, were added to the filter paper. Colony from tested organisms was transferred to the filter paper and rubbed onto the reagent with an applicator stick. The positive result was indicated by blue purple color formation within 10-15 seconds.

Molecular identification of Staphlococcus aureus

The detection of methicillin resistance is important not only in the clinical management and treatment of *S. aureus* infection (10), but also it was based on the amplification of the genes responsible for methicillin resistance and *S. aureus* genes and Coding toxins in diagnosing *Staphylococcus aureus* and distinguishing it from other species (11).

Results and discussion

The results presented in this chapter are based on 150 Atopic Dermatitis (AD) instances in total. Patients Suspected infected with *Staphylococcus aureus*, Dermatologists and criteria are used to confirm the clinical diagnosis of AD patients of Hanifan and Rajka (12), Total of 150 clinical samples were collected, only 54 sample (36 % isolates) were given the typical biochemical tests and characteristics of morphology that is specific to *staphylococcus aureus*, while the other 96 clinical (64 %) isolates belong to other species of staphylococci (8.6) or other pathogenic bacteria and fungi (33.3). Figure (1).

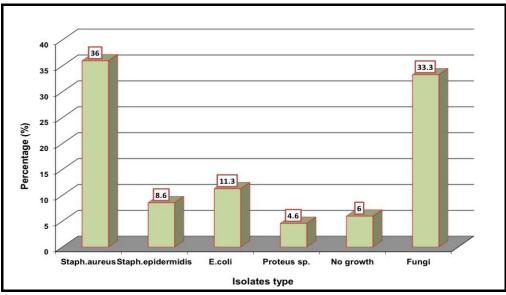


Figure (1): Bacterial types and fungi isolated from the skin of (AD) patients.

The Colonies of clinical specimens were cultured on blood agar media. The results found that bacteria have smooth, slightly and spherical shape. In addition, these colonies are able to form β hemolysis when grown on blood agar (13) (figure 2).

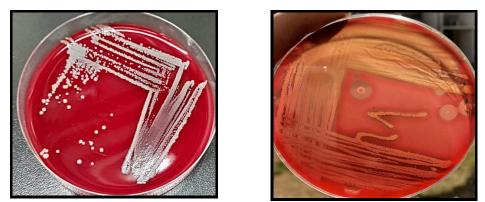


Figure (2): Growing of S. aureus colonies on blood agar at 37 °C for 24 hrs.

Fifty-four of clinical isolates form yellow (golden) colonies due to fermenting the mannitol salt change the phenol red to golden and resist high salts amount of MSA (Mannitol salt agar) selective medium. These gave the typical morphological characteristics of that specific to *S. aureus* (14,15). It was

a differential culture medium specific for *S. aureus* prevent the growth of other bacteria (figure 3) (16,17,18).



Figure (3): Growing of *S. aureus* colonies on MSA at 37 °C for 24 hrs.

Growth on chromagar, a medium specifically designed for the detection and isolation of *S. aureus*, was used to

further analyze these *S. aureus* isolates figure (4).

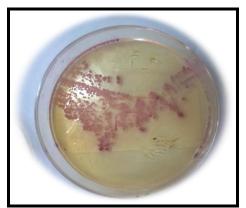


Figure (4): Staphylococcus aureus colonies on ChrmoagarTM.

The clinical specimens of diagnosed based examination of microscope. The Gram staining show the Staphylococci spp. properties when appear Gram positive reaction, nonspore-forming, arranged like irregular clusters and cocci shape under the microscope as shown in the Figure (5) (19).



Figure (5): Microscopic examination of *Staphylococci* isolates (10x).

Biochemical identification of *S. aureus* isolates

The basic biochemical tests were used to specimens shown a positive reaction for catalase, coagulase. But, it was a negative reaction for oxidase tests.

They are subjected to coagulase reactions that show the bacterial ability

to form coagulase enzyme which differentiates between S. aureus species (positive coagulase) and other Staphylococcus species (negative coagulase) due to reaction coagulase enzymes of bacteria with prothrombin of human blood and form staphylothrombin (clot) that convert the fibrinogen into fibrin Figure (6)(20).

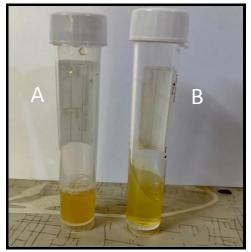


Figure (6): Coagulase test for *S. aureus*. A) negative result B) positive result.

Catalase was an enzyme that decomposes H2O2 (hydrogen peroxide) to oxygen and water, it prevented the toxic metabolites accumulation (21). The *S. aureus* isolates were positive Catalase genus differentiates from others *Streptococcus* genus (Figure 7). *S. aureus* were negative for oxidase, which differentiates them from others Micrococcus genus (22, 23) Figure (8).

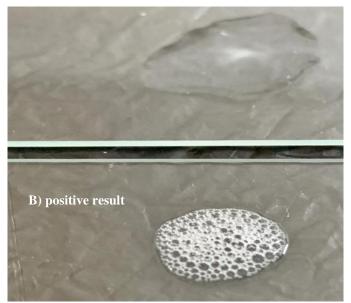


Figure (7): Catalase test for *S. aureus*.

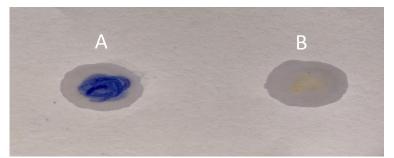


Figure (8): Oxidase test for S. aureus; A) Positive result B) Negative result.

Molecular identification of *Staphlococcus aureus*:

Detection of *mecA* gene in *S. aureus* isolates, the bacterial DNA amplified for this gene used PCR technique in a monoplex pattern by used specific primers, and the optimum condition to amplify this gene in PCR. The *mecA* gene confirmed by agarose gel electrophoresis and photographed under ultraviolet (UV) transilluminator shown in Figure (9)) were the amplification revealed a product of 147bp. The rate of clinical *S. aureus* isolates that gave a positive result for the *mecA* gene was 100%, The result consistent with the research of Jamil et al., (2017) (24) and Ghaznavi-Rad and Ekrami, (2018) (25) which was the rate of clinical *S. aureus* isolates. The one that gave a positive result for the *mecA* gene is 100% as well.

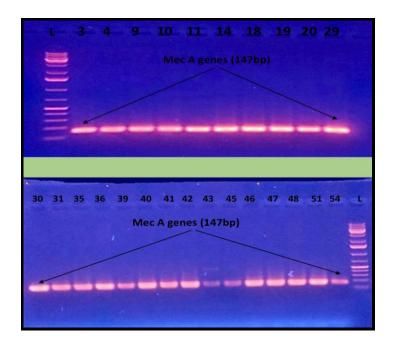


Figure (9): Gel electrophoresis of PCR products of mecA gene at 1%Agarose, 70 V and for 1 hr.

References:

- 1. Boguniewicz, M. and Leung, D.Y. (2010) Recent insights into atopic dermatitis and implications for management of infectious complications. Journal Allergy Clinical Immunology, 125: 4-13.
- Nowicka, D., & Nawrot, U. (2019). Contribution of Malassezia spp. to the development of atopic dermatitis. Mycoses, 62(7), 588-596.
- Orfali RL, Sato MN, Santos VG, Titz TO, Brito CA, Duarte AJ, Takaoka R, Aoki V. (2015)." *Staphylococcal enterotoxin* B induces specific IgG4 and IgE antibody serum levels in atopic dermatitis. International Journal Dermatology .54(8):898-904.
- Kong, H.H.; Oh, J.; Deming, C.; Conlan, S.; Grice, E.A. and Beatson, M.A. (2012). Temporal shifts in the skin microbiome associated with disease flares and treatment in children with atopic dermatitis. Genome Research .22:850e9.
- Nada, H.A.; Gomaa, N.I.; Elakhras, A.; Wasfy, R.; Baker, R.A. (2012); Skin colonization by superantigen-producing Staphylococcus aureus in Egyptian patients with atopic dermatitis and its relation to disease severity and serum

interleukin-4 level. International. Journal of Infectious Diseases. 16: e29-33.

- Prescott L.M. (2007). Laboratory Exercises in Microbiology. 7th ed. McGraw-Hill Higher Education. New York.
- 7. Bailey and Scott's (2002). Digngnostic Microbiology. 10th ed. Mosby. USA.
- 8. Brooks, G. F.; Carroll, K. C.; Butel, J. S.; and Morse, S. A. (2007). Jawetz, Melnick and Adelbergs Medical Inc. New York, 224-232.
- Harley, J.P., and Prescott L.M. (2007). Laboratory Exercises in Microbiology. 7th ed. McGraw-Hill Higher Education. New York.
- Francois, P.; Scherl, A.; Hochstrasser, D.; and Schrenzel, J. (2010). Proteomic approaches to study Staphylococcus aureus pathogenesis. Journal of proteomics, (73) (4):701-8.
- Wilson IG, Cooper JE, Gilmour A(1991). Detection of enterotoxigenic Staphylococcus aureusin dried skimmed milk: Use of polymerase chain reaction amplification and detection of Staphylococcal enterotoxin genes entB, entC1 and the thermonuclease gene nuc.

Applied. Environmental Microbiology .57: 1793-1789.

- 12. Hanifan, J.M. and Rajka, G. (1980). Diagnostic features of atopic dermatitis. Acta.Derma. Venereology. 92: 44-47.
- Gillespie, S. and Hawkey, P. M. (Eds.). (2006). Principles and practice of clinical bacteriology. John Wiley & Sons.
- Forbes, B.A.; Sahm, D. and Weissfeld, A. (2002). Baily and Scott's Diagnostic Microbiology, 11th edition, Saint Louis. Mosby Company
- Moura, S.; Gebreyes, W. A., Marques, M. F. S., Stipp, D. T. etal. (2018). Occurrence ofmethicillin-resistant Staphylococcus aureus and coagulase-negative staphylococci in dairy goat herds in Ohio, United States. Journal of Dairy Sciences. 101(9): 7804-7807.
- Sayed, M.; Soliman, A.; Elabd, H. and Abdallah, N. (2016). Detection of Methicillin-Resistant Staphylococcus aureus from Clinical Samples by CHROMagar TM and the in Vitro Effect of Topical Antiseptics. The Egyptian Journal of Medical Microbiology. 38(105):1-6.
- Faden, A. (2018). Methicillin-resistant Staphylococcus aureus (MRSA) screening of hospital dental clinic surfaces. Saudi Journal of Biological Sciences 4:2-22
- Aires-de-Sousa, M.; Rodrigues, S.; Conceiçao, T. and de Lencastre, H. (2018). Evaluation of different screening methodologies for the detection of methicillin-resistant *staphylococcus aureus* from environmental surfaces: Swabs,

Gauzes, and Polywipes. Microbial Drug Resistance Journal. 24(5): 585-589.

- Ho, J.; Boost, M. and O'Donoghue, M. (2015). Tracking sources of *Staphylococcus aureus* hand contamination in food handlers by spa typing, American Journal Infection Control. 43: 759-761.
- Brooks, G. F. (2010). Jawetz, Melnick, & Adelberg's medical microbiology/Geo. F. Brooks. New York; Chicago: McGraw Hill Medical. 30-75
- 21. Halt Jr, G. B. (2004). U.S. Patent No. 6,816,849. Washington, DC: U.S. Patent and Trademark Office.
- Becker, K.; Skov, R. and von Eiff, C. (2015). Staphylococcus, Micrococcus, and other catalase-positive cocci. In Manual of Clinical Microbiology, Eleventh Edition (354-382). American Society of Microbiology.
- 23. Jamil, S., Saad, U., & Hafiz, S. (2017). Can amoxicillin clavulanate be used for treating MRSA. Journal of Pharmacology and Medicinal Chemistry, 1(1), 21-23.
- 24. Ghaznavi-Rad, E. and Ekrami, A. (2018) Molecular characterization of methicillinresistant *Staphylococcus aureus* isolates, isolated from a burn hospital in Southwest Iran in 2006 and 2014. International journal of microbiology.
- 25. AbdulRazzaq, A., Shami, A. M., & Ghaima, K. K. (2022). Detection of vanA and vanB genes Among Vancomycin Resistant *Staphylococcus aureus* Isolated from Clinical Samples in Baghdad Hospitals. Iraqi Journal of Biotechnology, 21(1).