Molecular Detection of Cephalosporin Resistance Genes in *Escherichia coli* Isolated from Urinary Tract Infections in Baghdad Hospitals

Sally S. AL-Jubouri and AbdulMuhsin M. Shami

Institute of Genetic Engineering and Biotechnology for Postgraduate Studies, University of Baghdad

Received: 1/6/2022 Accepted: 21/8/2022 Published: December 20, 2022

Abstract: Escherichia coli is a normal flora in the human and animals gut, but also it is pathogenic in the patient with immune system disorder , is the leading cause of enteritis, urinary tract infection, septicemia and other Medical infections One hundred and eighty (180) midstream urine samples (MSU) were collected from patients with urinary tract infection (UTI) of all age and both sexes. Samples were collected from Al-Karma Teaching Hospital and Al-Yarmouk Teaching Hospital from the beginning of November 2021 until the end of February 2022. All isolates were diagnosed based on microscopic examinations and morphological characteristics using Eosin methylene blue (EMB), Blood agar and MacConkey agar. The diagnosis was confirmed through Vitek system and API E kit. Where the diagnostic results showed that E.coli had the highest percentage, representing 44.7%, followed by Klebsiella pneumonia with 16% and Pseudomonas aeruginosa. 8% and Proteus mirabilis 2.7%, Staphylococcus aureus 14.3%, Enterococcus spp, 6.3% and Fungi 8%. As for age, the age groups of (11-20) years had the highest rate of infection with 29.5%, followed by (21-30) years with a rate of 16.1, while the age groups over 60 and (1-10) were at 12.5% and (51-60) and (31-40) were 10.7%, and the lowest percentage of infection was (31-40), representing 8.0% .while was the injuries, 58% were in women, 42% in men Testing antibiotic sensitivity against 14 different antibiotics showed that E. coli was highest resistance Ceftriaxone (82%), Cefotaxime and Tetracyclic (78%), Cefixime (76%), Ceftazidime and Ampicillin (74%). While there is less resistance at Cefepime (68%) On the other hand, moderate resistance Cephalothin (50%), Cephalexin and Azetrenam (40%) The current study demonstrated that E.coli possessed a low-level resistance against Amikacin (34%), Ciprofloxacin (28%), Imipenem (16%), and Gentamycin (52%). In addition, PCR was used to identify cephalosporin resistance genes. It was concluded that all isolates were resistant to cephalosporin through the presence of the CTX-M-1 gene by (100%), while the results of the presence of the OXA-48 genes were (23.3%). Three resistant and three intermediate resistant isolates were selected for gene expression study.

Keywords: *OXA-48*, *CTX-M-1*, UTI, PCR, *E. coli*

Corresponding author: (Email: abdulmuhsin.m@ige.uobaghdad.edu.iq)

Introduction

Global population growth is accompanied by a wide spread of diseases caused by multidrug-resistant bacteria. Urinary tract infection (UTI) is the most common community-acquired and hospital-acquired bacterial infection typically characterized by a high rate of treatment failure and recurrence of

infection (1). Bacterial infection typically characterized by a high rate of treatment failure and recurrence of infection (2). They cause urinary tract infections, including *Escherichia coli*, *Staphylococcus saprophyticus*, *Klebsiella spp*, Proteus *spp*, and *Enterococcus* spp. (3).

The main pathogen responsible for infectious diseases in UTIs worldwide is the pathogenic *Escherichia coli* (UPEC). Urinary tract is successfully colonized by UPEC. UPEC isolates show different virulence factors that increase pathogenicity (4).

Escherichia coli is a Gram-negative bacteria, motile or non- motile a member of the Enterobacteriaceae, rod shaped, facultatively anaerobic, lactose fermentative. It is motile as it has peritrichous flagella. Most *E. coli* strains have the adhesive organs fimbriae (pili) that outspread from the bacterial surface into the surrounding medium (5,6).

It is employ several mechanisms of resistance agents different groups of antibiotics including; target modification, alteration of cell membrane permeability, production of enzymes, alteration of metabolic pathways and have the efflux pumps(7).

of One the most common mechanisms of resistance is the production of extended-release betalactamases (ESBL) and carbapmase that degrade all penicillins, cephalosporins, ximino, cephalosporins, and monomers (8).

Carbapenemase genes, mediated by elements mobile genetic carrying additional resistance elements, confer various resistance to groups antibiotics, resulting in multidrug resistance (MDR). OXA-48, a class D carbapenemase, is of major concern owing to its difficulty in detection and its association with treatment failure. *OXA-48* like Moreover. variants are plasmid coded and hence associated with rapid dissemination in community settings (9).

The CTX-M-type β -lactamases represent a group with a typical

extended-spectrum **B**-lactamase (ESBL)-resistance phenotype. These enzymes encoded by transferable plasmids. The enzyme responsible for this particular **ESBL** phenotype not affecting ceftazidime was named as CTX-M 1 in reference to its preferential hydrolytic activity against cefotaxime (CTX - M) (10).

Materials and methods Collection of samples

One hundred and eighty midstream urine samples (MSU) were collected from patients with urinary tract infection (UTI) of all ages and both sexes. Samples were collected from Al-Karama Teaching Hospital and Al-Yarmouk Teaching Hospital from the beginning. Identification of *E.coli* manual biochemical tests that were used Api20 E test. For final embedded in VITEK2 compact system.

Statistical analysis

The Statistical Analysis System- SAS (2012) program was used to detect the effect of different factors in study parameters. Least significant difference – LSD and (Analysis of Variation-ANOVA) was used to significantly compare between means. The Chi-square test was used to significant compare between percentage (0.05 and 0.01 probability) in this study.

Antimicrobial susceptibility testing

The E. coli isolates (50) were tested for antibiotic sensitivity by using different antibiotics. against 12 Amikacin, Imipenem, Azetrenam, Ampicillin, Cefepime, Cephalothin, Cefixime, Ceftriaxone, Ceftazidime, Cephale-xin Cefotaxime, and Ciprofloxacin (11).

Extraction and measured concentration and purity of DNA

Bacterial DNA extraction

DNA Extracted has been performed using (Qubit) NEB® (England) for all viteck-2 diagnosed *E.coli* isolates.

Detection of *E. coli 16SrRNA* gene using conventional PCR

Genes encoding the cephalosporin resistance determinants, OXA-48 and $CTX_{-}M_{-}I$, were investigated by PCR

using specific primers in table (1) of each primer reaction the following steps: an initial denaturation step at 94°C for 5 min; followed by 30 cycles of 94°C for 30 sec, 50°C for 45 sec and 72°C for 7 mins. Table (2) component of PCR master mix reaction. Where is the gel electrophoresis concentration 2% and voltage use 90 volt.

Table (1): Primer sequences used of PCR in this study

Primer	Primer sequence (5—3)	Product size bp	Reference	
16SrRNA	F- CTTAATCGACCATACGCTTTG	400	(12)	
	R- GTATTGTGTCTGCCATTAACC	400		
OXA-48	F -GCTGGTTCGCCCGTTTAA	159	(13)	
	R - ATTATCGGAATGCCAGCGGT	139		
CTX-M-1	F -GCTAAGCTCAGCCAGTGACA	733	(14)	
	R- CGCGGTGCTGAAGAAAGTG	755		

Table (2): Component of PCR Master Mix Reaction

PCR Master mix	x reaction components	Volume		
PC	R Premix	12.5Ml		
DNA	A template	5Ml		
Primers	Forward	1.5Ml		
Finners	Reverse	1.5μL		
nuclease-free H ₂ O		4.5μL		
Tot	al volume	25μL		

Results and discussion

One hundred and eighty midstream urine samples (MSU) were collected patients with from urinary infection (UTI) of all ages and both sexes. Samples were collected from Al-From 180 urine samples; preliminary results showed growth in 112 (62.3%) specimens and 68 samples no growth in the other (37.7) % E. coli had the highest percentage, representing 44.7%, followed by Klebsiella pneumonia with 16% and Pseudomonas aeruginosa. 8% Proteus mirabilis 2.7%, and Staphylococcus 14.3% aureus

Enterococcus spp 6.3% and fungi 8%, represents 8.0 % in table (3). Recent study shown similar result in Iraq by Al-Quraishi (2021) (15), the results were similar, as E.coli 68%, Klebsiella pneumonia 15%. Pseudomonas aeruginosa 10%, Proteus mirabilis 6%, Staphylococcus 65% and Enterococcus 35%. In Iraq a study by Khalil et al. (2016) (16) found that E. coli was the most frequent cause of UTIs in patients (28.6%), led by *Klebsiella spp.* (21.7%), and Staphylococcus spp. (16.7%), out of 143 total of urine samples.

Bacteria	No.	Percentage %
E.coli	50	44.7
Klebsiella pneumonia	18	16
Pseudomonas aeruginosa	9	8
Proteus mirabilis	3	2.7
Staphylococcus aureus	16	14.3
Enterococcus spp	7	6.3
Fungi	9	8.0
Total	112	100
Chi Square	-	85
p-value	-	< 0.001

Table (3): Numbers and percentage of total specimens (N=180)

As for age, the age groups of (11-20) years had the highest rate of infection with 29.5%, followed by (21-30) years with a rate of 16.1% in

table(4). While the age groups over 60 and (1-10) were at 12.5% and (51-60) and (31-40) were 10.7%, and the lowest percentage of infection was (31-40).

Table (4): Percentages of infection by age groups

Age group	Number infection	Percentage %
1-10	14	12.5
11-20	33	29.5
21-30	18	16.1
31-40	12	10.7
41-50	9	8.0
51-60	12	10.7
Over 60	14	12.5
Total	112	100
Chi Square	-	21.5
P –value	-	**0.001

Table (5): Percentages of infection by gender

Gender	Number	Percentage %			
Female	65	58			
Men	47	42			
Total	112	100			
Chi Square	=	2.5			
P –value	-	0.11			

Where it was found that the rate of infection in females was 58 % and in men 47% which represent. This study is similar to Saeed *et al.*, 2015 (17) study in Iraq, Erbil city, where the infection rate was (53.2% females, 46.8 % males) in table (5).

The antibiotic susceptibility test was carried out for *Escherichia coli* by Kirby Bauer disk diffusion method on a Mueller-Hinton agar according to. All

identified *E.coli* were exposed to antimicrobial agents in table (6).

Similar findings were reported in a study carried out in Iraq by AL-Taie(18) which showed that out of 80 isolates of *E. coli* were resistant at different rates Ciprofloxacin (47%), Ceftazidime (97%), Cefepime 72%, Ceftriaxone (72%), Ampicillin (100%), and Gentamycin (67%) Tetracycline (23%) and Imipenem (7%).

Antibiotic	Resistant		Intermediate		Sensitive		CI.	
(A.B)	Number Sample	%	Number Sample	%	Number Sample	%	Chi Square	P –value
Amikacin	12	24	21	42	17	34	4.88	0.08
Ampicillin	37	74	12	24	1	2	81.68	< 0.0001
Imipenem	8	16	12	24	30	60	32.96	< 0.0001
Azetrenam	20	40	22	44	8	16	13.76	0.001
Cephalothin	25	50	19	38	6	12	22.64	< 0.0001
Cefepime	34	68	16	32	0	0	12.76	< 0.0001
Cefixime	38	76	7	14	5	10	82.16	< 0.0001
Ceftriaxone	41	82	5	10	4	8	106.64	< 0.0001
Ceftazidime	37	74	12	24	1	2	81.68	< 0.0001
Cefotaxime	39	78	8	16	3	6	91.28	< 0.0001
Cephalexin	20	40	22	44	8	16	13.76	< 0.0001
Ciprofloxacin	14	28	10	20	26	52	16.64	< 0.0001
Gentamycin	26	52	19	38	5	10	27.44	< 0.0001
Tetracyclic	39	78	4	8	7	14	90.32	< 0.0001

Table (6): Percentages of antimicrobial susceptibility test for 50 E.coli isolates

Molecular identification of *E.coli* by *16S rRNA* gene, *OXA-48* and *CTX-M-1* gene

All Phenotypically positive isolates of *E.coli* strains were subjected to

molecular identification using the specific initiator of the *16S rRNA* gene to confirm its diagnosis by PCR. The results showed that all bacterial isolates belong to *E.coli* (Figure 1).

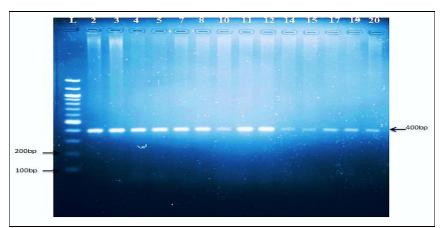


Figure (1): The gel electrophoresis of PCR product of 16SrRNA gene (400 bp) of E.coli, 100bp DNA ladder; (1% Agarose, 90 V for 70 mint).

The positive isolates identified by the Vitek 2 system assay gave a positive result in the detection of 16SrRNA gene by polymerase chain reaction amplification. All isolates were confirmed to be *E. coli* using 16SrRNA-

PCR. Included 30 (60%) *E. coli* were positive (100%).

The results of the current study agree with the results of the local study conducted by the researcher Ibrahim (2), where the percentage of bacterial isolates possessing 16 is 100%. The

current study also agrees with the study reached by the researcher Maleki *et al*. (19) in Iran.

The current results were similar to the study performed by Khalaf, *et al* (20) in Baghdad / Iraqi all 10 samples were identified as *E. coli* and it depends on the 16SrRNA gene 100%.

Detection of *E. coli CTX-M-1* gene using conventional PCR

To analyze the composition and molecular mechanisms of antibiotic resistance of Gram-negative bacteria - pathogens of urinary tract infections (UTIs) All the isolates were confirmed as *E. coli* using *CTX-M -1* PCR. Total 30 isolates included were positive

(100%). The positive result isolates are shown in figure (2) .AL -Nassyriah city in Iraq by Lhwak and Abbas (21). This study was conducted for detection of extended spectrum \(\beta \) lactamases enzyme and CTX-M-1 gene among E.coli causes urinary isolates that tract infections for pregnant women at gene CTX-M-1 was found in 17 isolates from 57 (29.8%)in E. Coli. Results were reported in a study by Ahmed *et al.* (22) the results of bacterial examination showed successful confirmation of E. coli which has been isolated from samples of, detection of CTX-M-1 gene was confirmed in 53.7% (36/67) of E. coli isolates.

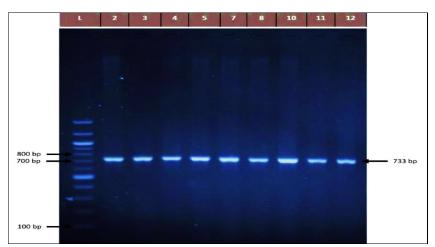


Figure (2): The gel electrophoresis of PCR product of CTX-M-1 gene (733 bp) of E.coli, 100bp DNA ladder; (1% Agarose, 90 V for 70 mints).

Detection of *E. coli OXA-48* genes using conventional PCR

The results showed in our current study, among the 30 isolates that were entered into the PCR machine, only isolates were found, which represent about 23.3%, containing the *OXA-48* gene. The positive result isolates are shown in figure (3). The current results were similar to the study performed in Algeria by Loucif *et al.*(23) The

genotyping results of carbapenemase among the obtained isolates are shown in of the 35 isolates, seven were carbapenemase producers, and four isolates were positive for were positive. bla OXA-48. However, a study conducted another study in Napal by Muktan et at (24). It has found that out of a total of 11 samples clinical that produce the enzyme carbapenemase, an isolate contains the OXA-48 gene.

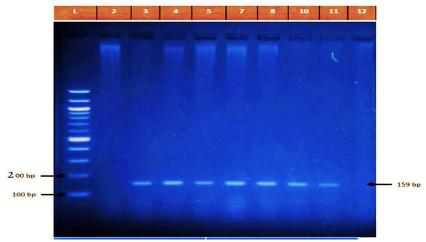


Figure (3): The gel electrophoresis of PCR product of *OXA-48 gene* (159 bp) of *E.coli*, 100bp DNA ladder; (1% Agarose, 90 V for 70 mints).

Conclusions

The current study showed that most *E. coli* isolates have a high rate of antibiotic resistance. Ceftazidime is an effective cephalosporin antibiotic used in the treatment of urinary tract infection caused by *Escherichia coli*. Identification of isolates containing *OXA-48* and *CTX-M-1* genes using conventional PCR. These samples are resistant to cephalosporin groups when compared to the molecular and phonotypical susceptibility test.

References

- Kemaa, A. A.; Al-Ugaili, D, N. and Al-Azzawi, I. S. (2019). Multilocus Sequence Typing of Multi-Drug Resistant *Escherichia* coli isolated from Urinary Tract Infections. International Journal of Pharmaceutical Research, 11(3).16-28.
- Islam, M. R.; Hoque, M. J.; Uddin, M. N.; Dewan, A.; Haque, N. B.; Islam, M. T. and Hasan, M. A. (2022). Antimicrobial Resistance of *E.coli* Causing Urinary Tract Infection in Bangladesh. Mymensingh Medical Journal, 31(1), 180-185.7
- 3. Masinde, A.; Gumodoka, B.; Kilonzo, A.; & Mshana, S. E. (2009). Prevalence of urinary tract infection among pregnant women at Bugando Medical Centre, Mwanza, Tanzania. Tanzania Journal of Health Research, (3). 11-15

- Schmider, J., Bühler, N., Mkwatta, H., Lechleiter, A., Mlaganile, T., Utzinger, J. & Becker (2022). Microbiological Characterisation of Community-Acquired Urinary Tract Infections in Bagamoyo, Tanzania: A Prospective Study. Tropical Medicine and Infectious Disease, 7(6), 100 -123.
- Lillington, J.; Geibel, S.; & Waksman, G. (2014). Biogenesis and adhesion of type 1 and P pili. *Biochimica Biophysica Acta* (BBA) General Subjects, 1840(9), 2783-2793.
- 6. Jaweetz E, Melnick J A and Adelberg E A. (2016). Review of Medical Microbiology 27th ed. McGraw. Hill Higher Education 850.10-11
- 7. Kapoor, G., Saigal, S., & Elongavan, A. (2017). Action and resistance mechanisms of antibiotics: A guide for clinicians. Journal of anaesthesiology, Clinical Pharmacology, (3), 30-33.
- 8. Roshani, M.; Goudarzi, H.; Hashemi, A.; Ardebili, A.; Erfanimanesh, S.; & Bahramian, A. (2018). Detection of IS903, IS26 and ISEcp1 elements in *CTX-M* producing *Klebsiella pneumoniae* and *Escherichia coli* isolates from patients with leukemia in Iran. Jundishapur Journal of Microbiology, 11(12). 160-180
- 9. Oueslati, S.; Retailleau, P.; Marchini, L.; Dortet, L.; Bonnin, R. A.; Iorga, B. I.; & Naas, T. (2019). Biochemical and structural characterization of *OXA-405*, an *OXA-48* variant with extended-spectrum β-lactamase activity. Journal of Microorganisms, 8(1), 24-30

- Cantón, R.; González-Alba, J. M.; and Galán, J. C. (2012). CTX-M Enzymes: Origin and Diffusion. Frontiers in Microbiology, (3), 110-112.
- 11. AbdulRazzaq, A. B.; Shami, A. M. and Ghaima, K. K. (2022). Detection of *vanA* and *vanB* genes Among Vancomycin Resistant *Staphylococcus aureus* Isolated from Clinical Samples in Baghdad Hospitals. Iraqi Journal of Biotechnology. 21(1): 19-25...
- 12. Al-Saadi, Z. H. A., & Abdullah, R. M. (2019). phenotypic and molecular detection of *Escherichia coli* efflux pumps from uti patients. *Biochemical and Cellular Archives*, 19(Suppl. 1), 2371-2376.
- Hasoon, N. A., & Hamed, S. L. (2019). Molecular Characterization of Carbapenemase-Producing Gram-negative Bacteria Isolated from Clinical Specimens in Baghdad, Iraq. *Iraq*. Journal of Microbiology.13 (2), 1031-40.
- Auda, I. G., Al-Kakei, S. N. H., & Hamed, S. L. (2013). Occurrence of CTX-MI Type β-lactamases Gene in Certain Gram Negative Bacteria. Iraqi Postgraduate Medical Journal, 12(2).3-4
- 15. Al-Quraishi, R., Abud –kadim. (2021). Virulence Evaluation of Uropathogenic Escherichia coli Strains According to Molecular Screening of The Toxins Genes (sat, vat, hlyA and cnf1). Baghdad University Thesis
- Khalil, Z. K. (2016). Uropathogenic Infections among Patients with Diabetes in Baghdad City. Iraqi Journal of Science. 57,360–366.
- Saeed, C. H., AL-Otraqchi, K. I., & Mansoor, I. Y. (2015). Prevalence of urinary tract infections and antibiotics susceptibility pattern among infants and young children in Erbil city. Zanco Journal of Medical Sciences, 19(1), 915_922-915_922.
- 18. AL-Taie, S. F. (2021). The Synergistic Effects of Green Synthesized ZnO

- Nanoparticles and Ceftriaxone on Uropathogenic. Al-Nahrein University Thesis.
- Maleki, D.; Jahromy, S. H.; Karizi, S. Z. and Eslami, P. (2017). The Prevalence of acrA and acrB Genes among Multiple-Drug Resistant Uropathogenic Escherichia coli Isolated from Patients with UTI in Milad Hospital, Journal of Pharmacy and Pharmaceutical Sciences. (11), 53:57.
- 20. Khalaf, S. A.; Mahmood, N. N.; and Saleh, M. A. D. (2021, September). Detection of tetM, armA, blaPER-1 and blaIMP genes in *E. coli* isolates among the gram negative bacteria that cause urinary tract infections. Journal of Physics Conference. 1999,12-21.
- Lhwak, N. S.; & Abbas, Y. A. (2018).
 Detection of Extended Spectrum β– Lactmase Genee CTX-M -1 in Escherchia coli and Klebseilla pneumonia Isolated from Urinary Tract Infection of Pregnant Women in Al-Nassyriah City. University of Thi-Qar Journal of Science, 2(4), 92-96.
- 22. Ahmed, I. M.; Aldabbagh, S. Y.; & Jwher, D. M. (2021). Molecular characterization of extended spectrum cephalosporin resistant *Escherichia coli* isolated from dogs. Iraqi Journal of Veterinary Sciences, *35*(3), 473-478
- 23. Loucif, L.; Chelaghma, W.; Bendjama, E.; Cherak, Z.; Khellaf, M.;Khemri, A.; & Rolain, J. M. (2022). Detection of blaOXA-48 and mcr-1 Genes in Escherichia coli Isolates from Pigeon (Columba livia) in Algeria. Journal Microorganisms, 10(5), 975.
- 24. Muktan, B.; Thapa Shrestha, U.; Dhungel, B.; Mishra, B. C.; Shrestha, N.; Adhikari, N. & Ghimire, P. (2020). Plasmid mediated colistin resistant mcr-1 and co-existence of *OXA-48* among *Escherichia coli* from clinical and poultry isolates: Nepal. Journal Gut Pathogens, 12(1), 1-9.