

Investigation of Some Carbapenemase Resistance Genes of *Klebsiella pneumoniae* Isolates and their Role in the Antibiotics Resistance

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Abstract: Klebsiella pneumoniae is ubiquitous gram-negative encapsulated pathogen that has long been associated with variety of infections. The aim of the current study was to determine the presence of carbapenemase genes in K. pneumoniae isolates obtained from different sources and their role in the antibiotics resistance. A total of 210 clinical samples were collected from patients of both sexes ranging between 10 to 80 years old from different sources; (urine, sputum, wound swabs, blood, and liver abscess) who admitted from different hospitals in Baghdad from (March, 2022 to July, 2022). Results appeared that only 90 isolates were identified as K. pneumoniae based on differential and selective media, biochemical tests followed by Vitek 2 systems and molecular detection using 16S rRNA. Antibiotic susceptibility test of K. pneumoniae isolates against 13 antibiotics were established using disc-diffusion technique. Result appeared that all isolates were 100% resistant to Vancomycin. While, Colistin had the lowest resistance rate 25.55% manifested that was the most effective antibiotic. Carbapenemase genes; New Delhi metallolactamase NDM-1, Verona integron-encoded metallo β -lactamase VIM and imipenemresistant carbapenemases IMP were detected using polymerase chain reaction PCR technique. It was concluded that antibiotics resistance rate was high in K. pneumonia isolates. Moreover, Colistin recommended for the treatments of carbapenemase-producing K. pneumonia. The PCR results revealed that IMP gene was not detected, while the percentage of NDM-1 and VIM genes were 23% and 3% respectively.

Keywords: Antibiotics resistance, carbapemase genes, K. pneumoniae, polymerase chain reaction.

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Introduction

Antimicrobial resistance is one of the most significant issues increased hospitals. infection in The Κ. pneumoniae accountable among onethird of all Gram-negative infections in the hospitals (1). Furthermore, that makes it the second most significant opportunistic Enterobacterium in nosocomial and society infections followed Escherichia coli involved pneumonia, meningitis, liver abscess, UTIs and others. Moreover, bacteria can even result in high rates of illness and mortality when treated improperly (2). K. pneumoniae isolates can The develop several mechanisms that results antibiotics resistance include; in Carbapenem resistance due to production of carbapenemases, the overuse of carbapenems antibiotics and the horizontal gene transfer of plasmids carry on multi resistance genes would be the reason of carbapenems to resistance and leads to limitations in the treatment options against these drugresistant bacteria (3,4). Other mechanisms such as alteration outer

membrane permeability, ESBL, over expression of AmpC that hydrolyze many beta-lactam rings of antibiotics, mutations of porins and penicillinbinding proteins (PBP), alteration of metabolic pathways, the presence of efflux pumps and biofilm formation capacity (5).

According to the Ambler which on classification depends sequence and structural homology, class B/ metallo- β -lactamases that hydrolyze carbapenem in a zinc dependent manner involved: imipenem-resistant carbapenemases IMP, Verona integronencoded metallo β -lactamase VIM and New Delhi metallolactamase NDM. They are encoded by *blaIMP*, *blaVIM* and *blaNDM*, respectively (6). The NDM has been detected in carbapenemresistant K. pneumoniae as well as in E. coli in a patient who has been relocated in 2009 from India (7). Multi-drug resistant Enterobacteriaceae carrying NDM-1 gene has been recognized worldwide, bacteria with NMD-1 may be responsible and more dissemination for urinary tract infections (8). Otherwise, the VIM and IMP families share certain similarities and both are susceptible to all β -lactam inhibitors and hydrolyze all β -lactams except monobactams (9). Hence, aim of this study: isolation and identification of K. pneumoniae isolates from different clinical sources as well as determination of multi-drug resistant (MDR) isolates susceptibility using antibiotic test. molecular detection of some carbapenemase genes including; NDM-1. IMP and VIM.

Materials and methods

Samples collection and bacterial identification

The project was approved by Baghdad College of Science's ethics committee (CSEC/0523/0042). In this study two hundred ten clinical samples were obtaining from patients including both sex, their age ranging from (10 to 80) years with different sources (urine, sputum, blood, wound swabs and liver abscess) who admitted from different hospitals in Baghdad (Central Children and Medical city / Educational lab, Teaching Laboratories /Medical city, Al-Yarmuk Hospital, Al-Karama teaching hospital and Al - Kadhimiya hospital), throughout the period from March, 2022 to July 2022. All isolates were identified using selective and differential culture media, biochemical tests, followed by Vitek 2 system and molecular detection using the 16S rRNA gene.

Antibiotics susceptibility of *K*. *pneumoniae* isolates

Disk -diffusion method usually test the susceptibility of thirteen antibiotics disc. Disks containing; Ampicillin AMP (10 µg), Cefixime CFM (5 µg), Cefotaxime CTX (30 µg), Chloramphenicol CHL (30 μg), Dorepenem DOR (10 µg), Imipenem IMP (10µg), Nalidixic acid NAL (30 (30 Tetracycline μg), TET μg), Vancomycin VA (30µg), Streptomycin STR (10 µg), Nitrofurantoin NIT (300 Amoxicillin-clavulanate AMC μg), $(10/20 \mu g)$. Otherwise, Colistin CST (10 µg) was examined using the minimum inhibitory concentration (MIC) by the broth microdilution method. All disks were obtained from (Liofilchmem) Italy), after the period of incubation at 37ºC. The results were interpreted in accordance with the Clinical Laboratory Standards Institute.

DNA extraction

Extraction of genomic DNA using Monarch® Genomic DNA Purification Kit (NEB, England). The concentration of the extracted DNA was examined using Qubit 4.0 (Invitrogen/USA).

Polymerase chain reaction (PCR) Primers design

The specific primers of *NDM-1*, *IMP* and *VIM* genes of *K. pneumoniae* isolates were check for the right annealing temperature using software called Geneious Prime bioinformatics. The primers were further evaluated using a variety of internet applications, including Oligo Analyzer by IDT (Integrated DNA Technology). The primers sequences were showed in (Table 1). Traditional PCR technique started with denaturation phase at 95 °C/ 5 minutes, followed by 30 cycles of denaturation at 95 °C. After that reaction mixture annealed at at 35 cycles for 55 C/ 45 seconds. The reaction mixtures are then subjected to 35 cycles of the extension stage over the course of one minute. The amplification ultimately ceases after seven minutes of the final extension stage at 72 °C. Then PCR product have been visualized using an agarose gel (2%) containing RedSafe dye and an ultraviolet (UV) light.

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Primer name		Sequence (5´-3´)	Ambilicon size(bp)	References
16S RNA	F	CGGTCTGTCAAGTCGGATGT	172	This study
	R	AGCGTCAGTCTTTGTCCAGG	172	
IMP	F	ATTCTCAATCCATCCCACGT	150	This study
	R	TGTGTCCTGGGCCTGGATAA	150	
VIM	F	TCATGGCTATTGCGAGTCCG	127	This study
	R	GCGATATGCGACCAAACACC	127	
NDM-1	F	GCATTAGCCGCTGCATTGAT	146	This study
	R	TAGGAAGTGTGCTGCCAGAC	140	

Table (1): The Designed primers sequences utilized in this study

Statistical analysis

The Statistical Analysis System SAS program (10) was employed to determine the impact of various factors on study's parameters. The Least Significant Difference (LSD) test (Analysis of Variation, ANOVA) was used in order to considerably compare between the means. In this study, the Chi-square test was used to compare percentages with p<0.05.

Results and discussion

Two hundred ten clinical samples were collected from various sources (urine, sputum, blood, wound swabs and liver abscess). Samples were transferred to the lab and streaked on sterile Macconkey and CHROMagar media after samples incubation for 24 hrs. Only 90 isolates showed pink and mucoid colonies on Macconkey agar due to lactose fermentation ability and metallic blue color on CHROMagar

Orientation medium which is define as a chromogenic, differential, and highly selective media for detecting Κ. in remarkably pneumoniae green colonies which was agreement with (11) as mentioned in (Figure 1). Biochemical tests also showed positive result for Urease, Citrate, Catalase and Vogues-Proskauer. Otherwise, negative for Indole, Oxidase and Methyl Red. Vitek 2 System results were utilized to confirm the identity of K. pneumoniae isolates. Distribution of K. pneumoniae isolates were illustrated in (Table 2), which represented that UTIs (57.7%) and sputum (24.4 %) have the highest percentage value, followed by blood (10%), wound swabs (5.5%) and liver abscess that represented the lowest percentage (2.2%). This is because of the UTI а relatively prevalent justification in current practice for consultation and antibiotic prescription.

Results agreement with local study of high percentage for urine 42.26% followed by sputum 26.70% (12). Furthermore, result was consistent with the findings of Tehran hospital and found that *K. pneumoniae* was obtained from different sources including; urine, sputum, and blood with percentages of 61.7%, 11.7%, and 8.5%, respectively (13).



Figure (1): Colonies of *K. pneumoniae* on: A: Macconkey agar, B: CHROMagar after 24 hours of incubation at 37 C.

Types of Samples	Total Samples N.O	Bacterial strain N.O	Percentage (%*)	Percentage (%**)		
Urine	105	52	50%	57.7%		
Sputum	41	22	19.5%	24.4 %		
Wound	31	5	14.7%	5.5%		
Blood	27	9	12.8%	10%		
Liver abscess	6	2	2.8%	2.2%		
Total	210	90	100%	100%		
P-value			0.0001 **	0.0001 **		
** (P≤0.01).						

 Table (2): The number and percentage of K. pneumoniae isolated from different sources.

Percentage (%*) distribution of *K. pneumoniae* relative to the samples, Percentage (%**) distribution of *K. pneumoniae* relative to the overall no. of *K. pneumoniae* strain.

Antibiotic susceptibility test of *K. pneumoniae*

Disk diffusion method was employed for antibiotics susceptibility of 90 K. pneumoniae isolates against antibiotics thirteen including; (Ampicillin, Cefixime. Cefotaxime. Chloramphenicol, Colistin, Dorepenem, Imipenem, Nalidixic acid, Tetracycline, Vancomycin, Streptomycin, Nitrofurantoin. Amoxicillinclavulanate). mentioned as in (Figure 2). The emergence of antimicrobial drug resistant strains has been reported in K. pneumonia isolated

from community and hospital acquired infections (14). The results revealed that high resistance of isolates against Vancomycin (100%), Ampicillin (97.77%), Amoxicillin-clavulanate (96.66%) followed by Cefixime (94.44%) and Cefotaxime (93.33%) these results are confirmed with a local study in Al-kufa, Iraq (15). On the other hands, isolates showed (35.33%) for Nalidixic acid and (25.55%) for Colistin. From the present study colistin represented high sensitivity that were approval with a study by Abdelhamid et al. (16) who notified that colistin was effective against all 50 Egyptian isolates that were examined. While, other shown resistance isolates to Nitrofurantoin (66.66%), Tetracycline Streptomycin (53.33%), (56.66%),Chloramphenicol (41.11%). production of specified enzymes, reduced cell permeability due to Omps depletion and over expression of efflux pumps are the main mechanisms that contribute to antibiotic resistance. Carbapenemase play an important role in MDR resistant mechanism which hydrolyze β -lactam antibiotics (17).

Colistin has become more widely used to treat Gram-negative bacteria that are resistant to carbapenem and multidrug resistant (MDR) as a rescue therapy, either by itself or in conjunction with one or more antimicrobials (18). The mechanism of action of colistin is based on the electrostatic interaction between the cationic residues of the drug's diaminobutyric acid, and the anionic phosphate groups of lipid A presence in bacterial lipopolysaccharide (LPS). It destabilizes the LPS, resulting in raised cell membrane permeability and causes cell leakage and death. So it has been used as a first-choice medication, particularly in critical care units. due to high rates of carbapenemresistance among KPCproducing *K. pneumonia* (19).



Figure (2): Antibiotics sensitivity test of 90 K. pneumoniae isolates obtained from various sources.

Molecular detection of *K. pneumoniae* using *16S rRNA*

The *16S rRNA* gene, a gene used to identify *K. pneumoniae*, has been tested using a polymerase chain reaction on each extracted DNA sample. The results of the PCR revealed positive *16S rRNA* gene which appeared in single band with molecular base 172 bp by comparing the molecular size of the bands to a 100bp DNA ladder

(Figure3). The findings presented here are in agreement with previous study by Ghaima and Tamara (20), who collected 260 clinical samples from urinary tract infection patients, and the results showed that only 76 of the 260 bacterial isolates belonged to *K. pneumoniae* by combining traditional methods with a molecular diagnostic approach based on the *16S rRNA* gene (159 bp).



Figure (3): Gel electrophoresis of 16S rRNA gene (172bp) amplification for K. pneumoniae. Lane L: 100-200 bp DNA ladder; Lanes 1-15: K. pneumoniae isolates positive. (2% agarose gels and stained with Red safe, 80V for 80 min.).

Molecular detection of *NDM-1*, *VIM*, *IMP* genes

According to multiple drug resistance (MDR) of *K. pneumoniae* a totally of 80 isolates were amplified with PCR for diagnosis the presence of *NDM-1*, *IMP* and *VIM* genes using specific designed primers.

The results of DNA extraction by gel electrophoresis were PCR-

positive for *NDM-1* (146bp) and *VIM* (127bp) as compared with 100bp DNA Ladder as illustrated in (Figures 4, 5). Otherwise, *IMP* (150bp) gene was not detected. Results revealed that *NDM-1* was the most prevalent of Carbapenemase detect genes with (18/23%) *Klebsiella* isolates. While, *VIM* gene presented only in two (3%) isolates.



Figure (4): Agarose gel electrophoresis of PCR for the *NDM-1* gene (146bp). Lane M: 100-200 bp DNA ladder; Lanes 1-9: *K. pneumoniae* isolates. (agarose 2%, and stained with Red safe, 80V for 80 min.).



Figure (5): Agarose gel electrophoresis of PCR for the VIM gene (127bp). Lane M: 100-200 bp DNA ladder; Lanes 1-9: K. pneumoniae isolates. (agarose 2%, with Red safe stain, 80V for 80 min.).

The enzymes of the VIM, IMP, and NDM are groups in Ambler class B, that cannot be physically bound to the beta-lactam substrate and therefore escape the action of beta-lactamase inhibitors such as clavulanic acid and sulbactam and are able to bind to all beta-lactam classes, except monobactam (21). These genes responsible for the production of carbapenemase and carbapenems resistance *K*. in pneumonia. Emerging NDM-1 provides resistance to all β -lactam antibiotics; it primarily seen in E. coli, K. is pneumoniae, and A. baumannii as hosts. It has been determined that Pakistan and India are reservoirs of NDM producers due to their NDM-1 gene (22). The NDM gene which was found in 43% of the isolates in the Egyptian study may be the causes of carbapenem resistance, according to PCR data, this finding may be strongly linked to a number of resistance mechanisms that resulted in resistance to the majority of antibiotic (23). Study from Turkey classes examined 77 isolate of K. pneumoniae and revealed that nine isolates (10.84%)

produced *NDM-1*(24). Other carbapenems genes; VIM and IMP were detected primarily in Pseudomonas aeruginosa and commonly observed at shallow rate among a Enterobacteriaceae. On the other hands, An Iraqi study of K. pneumoniae strains reminded no blaVIM gene distribution among carbapenemresistant K. pneumonia and just five strains were illustrated as positive for blaIMP gene 5 (9.1%), while results showed the presence of a *blaNDM-1* gene in 37 (67.27%) carbapenemresistant K. pneumoniae strains (25). Moreover, Mahmood (26) who revealed that Carbapenemase-producing Е. has been cloacae reported carbapenemase genes blaNDM (n=4/8, 50%) and *blavim* (n=3/8, 37%).

Conclusion

In this study, all of *K*. *pneumoniae* isolates revealed 100% resistant to Vancomycin followed by Ampicillin, Amoxicillin-clavulanate, Cefixime. On the other hands the lowest resistance was against colistin with a percentage 25.55% that was the most

effective antibiotic applied in the present study. Molecular detection of carbapenemase genes achieved using PCR technique. The distribution of carbapenemase *NDM-1* gene was more frequent gene than *VIM*. On the other hands, *IMP* gene not detectable. All *K. pneumoniae* isolates that have carbapenemase genes are resistant to most antibiotics used.

References

- 1. Liu, W.; Tang, J.; Lyu, J.; Wang, J.; Pan, Y.; Shi, X. and Wang, L. (2022). Discrimination between carbapenem-resistant and carbapenem-sensitive *Klebsiella pneumoniae* strains through computational analysis of surface-enhanced Raman spectra: a pilot study. Microbiology spectrum, 10(1): e02409-21.
- Rhumaid, A. and Al-Mathkhury, H. (2015). Detection of *blaKPC* Gene in Some Clinical *Klebsiella pneumoniae* Isolates in Baghdad. Iraqi Journal of Science, 56(4A): 2853-2861.
- AL-Jubouri, S. and Shami, A. (2022). Molecular Detection of Cephalosporin Resistance Genes in *Escherichia coli* Isolated from Urinary Tract Infections in Baghdad Hospitals. Iraqi Journal of Biotechnology, 21(2): 145-152.
- Arcari, G.; Polani, R.; Bruno, F.; Capitani, V.; Sacco, F.; Menichincheri, G., *et al.* (2023). Ceftazidime–avibactam resistance in *Klebsiella pneumoniae* sequence type 37: a decade of persistence and concealed evolution. Microbial Genomics, 9(2): mgen000931.
- Rahal, B.; Salman, A. and Mohamed, K. (2021). The Role of EDTA in Biofilm Eradication of *Klebsiella pneumoniae* Isolated from Wound Infections. Iraqi Journal of biotechnology, 1(20): 96-102.
- Bush, K. and Bradford, P. (2020). Epidemiology of β-lactamase-producing pathogens. Clinical Microbiology Reviews, 33(2): 10-1128.
- Yong, D.; Toleman, M.; Giske, C.; Cho, H.; Sundman, K.; Lee, K., *et al.* (2009). Characterization of a new metallo-βlactamase gene, *bla NDM-1*, and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. Antimicrobial Agents and Chemotherapy, 53(12): 5046-5054.

- Abbas, F. (2022). Prevalence of Waterborne bla NDM-1 Gene Producing Carbapenemresistant Klebsiella pneumoniae from Al-Hillah River Water, Babylon Province, Iraq. Journal of Pure and Applied Microbiology, 16(3):1873-1877.
- 9. Abou-assy, R.; Aly, M.; Amasha, R.; Jastaniah, S.; Alammari, F. and Shamrani, M. (2023).Carbapenem Resistance Mechanisms. Carbapenemase Genes Dissemination, and Laboratory Detection Methods: A Review. International Journal of Pharmaceutical Research and Allied Sciences, 12(1): 123-138.
- 10. Stokes, M.; Davis, C. and Koch, G. (2012). Categorical data analysis using SAS. SAS institute.
- 11. Omar, F. and Ibrahim, A. (2023). The prevalence of Integron class I and II among multi-drug resistance producing *Klebsiella pneumoniae*. Iraqi Journal of Agricultural Science, 54(3): 619-629.
- 12. Al-Ruobayiee, M. and Ibrahim, A. (2023). The Relationship between OqxAB Efflux Pump and Drug Resistance in *Klebsiella pneumoniae* Isolated from Clinical Sources. Al-Rafidain Journal of Medical Sciences, 1(5): S106-112.
- Seifi, K.; Kazemian, H.; Heidari, H.; Rezagholizadeh, F.; Saee, Y.; Shirvani, F., *et al.* (2016). Evaluation of biofilm formation among *Klebsiella pneumoniae* isolates and molecular characterization by ERIC-PCR. Jundishapur Journal of microbiology, 9(1): e30682.
- 14. Shabaa, R. (2014). Detection of *CTX-M-1* gene Among *Klebsiella pneumonia* Isolates in An Najaf Province. Iraqi Journal of Biotechnology, 13(2): 128-133.
- 15. Al-Hasnawi, H. (2020). Frequency of Carbapenem-Resistant *Klebisella pneumoniae* from Najaf Hospitals and Investigate the Dissemination of Class 1 Integron among Isolates. Phd Thesis. Collage of Medicine, AlKufa University.
- 16. Abdelhamid, S.; Abd-Elaal, H.; Matareed, M. and Baraka, K. (2020). Genotyping and Virulence Analysis of Drug Resistant Clinical *Klebsiella pneumoniae* isolates in Egypt. Journal of Pure and Applied Microbiology, 14(3): 1967-1975.
- 17. Logan, L. and Weinstein, R. (2017). The epidemiology of carbapenem-resistant Enterobacteriaceae: the impact and evolution of a global menace. The Journal of infectious diseases, 215(suppl_1): S28-S36.

- 18. Lim, L.; Ly, N.; Anderson, D.; Yang, J.; Macander, L.; Jarkowski, A., *et al.* (2010). Resurgence of colistin: a review of resistance, toxicity, pharmacodynamics, and dosing. Pharmacotherapy. The Journal of Human Pharmacology and Drug Therapy, 30(12): 1279-1291.
- 19. Kadum, S. and Al Rubaeye, D. (2020). Colistin Susceptibility in Carbapenem Resistant *Klebsiella Pneumoniae* and their Ability of Biofilm Formation. Iraqi Journal of Science, 61 (3): 517-527.
- 20. Ghaima, T. M. (2022). Molecular Detection of acrAB and oqxAB Genes in Klebsiella pneumoniae and Evaluation the Effect of Berberine on their Gene Expression. Iraqi Journal of Biotechnology, 21(2): 124-135.
- 21. Bush, K. (2018). Past and present perspectives on β-lactamases. Antimicrobial Agents and Chemotherapy, 62(10): 10-1128.
- 22. Hamoodi, D. (2022). Detection of Plasmid-Mediated *blaNDM1* and *blaNDM2* Genes in Clinical Isolates of *Acinetobacter baumannii*

from Iraqi Patients. Systematic Reviews in Pharmacy, 13(3): 342-348.

- 23. Hamad, S. (2022). Prevalence of Carbapenemase Genes in *Klebsiella pneumoniae* Isolates from Patients with Urinary Tract Infections in Baghdad Hospitals. Iraqi Journal of Biotechnology, 21(1): 102-114.
- 24. Yıldız, S.; Kaşkatepe, B.; Avcıküçük, H. and Öztürk, Ş. (2017). Performance of CarbaNP and CIM tests in OXA-48 carbapenemaseproducing Enterobacteriaceae. Acta Microbiologica et Immunologica Hungarica, 64(1): 9-16.
- 25. Hussein, N. (2018). Emergence of *NDM-1* among carbapenem-resistant *Klebsiella pneumoniae* in Iraqi Hospitals. Acta Microbiologica et Immunologica Hungarica, 65(2): 211-227.
- Mahmood, S. (2022). The prevalence of blaNDM, blaVIM genes among Enterobacter cloacae bacteria. Iraqi Journal of Agricultural Sciences, 53(4): 958-964.